# Calibration System and Calibration Method of Biochip Temperature Sensors

Sang-Yoon Kim<sup>1,3</sup>, Jong-Dae Kim<sup>2,3</sup>, Yu-Seop Kim<sup>2,3</sup> Hye-Jeong Song<sup>2,3</sup> and Chan-Young Park<sup>2,3</sup>\*

<sup>1</sup>Department of Computer Engineering, Hallym University, Korea
 <sup>2</sup>Department of Ubiquitous Computing, Hallym University, Korea
 <sup>3</sup>Bio-IT Research Center, Hallym University, Korea
 { lastamor, kimjd, yskim01, hjsong, cypark}@hallym.ac.kr

#### Abstract

Currently, a microanalysis technology is employed in detection systems in fields such as medical science, food safety, and environmental monitoring, and it is being developed in the form of a lab-on-a-chip. However, the reproducibility and sensitivity of biochips are essential factors for its commercialization. Because this requires accurate temperature control, there should be no errors in the temperature sensor used for the biochip. This paper proposes a system that calibrates the temperature sensor of the biochip by directly heating the biochip and using the difference between the measured values of the temperature sensor attached to the biochip and the precision temperature sensor on the biochip surface. In addition, directly heating the biochip was observed to shorten the time taken to reach the target temperature, compared to the chip calibration by placing it in the constant-temperature water bath. The shortened calibration time of the biochip enables us to realize reduced production cost when the chips are manufactured on a large scale.

Keywords: Biochip, Temperature calibration, Temperature sensor

### 1. Introduction

Developed in 1983 by Kary Mullis[1, 2], PCR is now a common and often indispensable technique used in medical and biological research labs for a variety of applications[3, 4].

Biochips can obtain information regarding multiple genes within a short period of time, and this enables the analysis of the gene. Further, biochips are used to diagnose various diseases based on reactions with components inside the body of a target person to be examined.

Biochips are divided into microarray and microfluidics chip depending on the form[5]. The commercialization of biochips requires them to be reliable in terms of their reproducibility and sensitivity, and therefore, accurate temperature control and calibration of the biochip's temperature sensor are required. To realize temperature control, a heater and a temperature sensor are usually attached to the surface on which the DNA is amplified [6-10]. For this, the biochip is equipped with various temperature sensors such as a metal film, a thermistor, and a thermocouple; however, a calibration method has not yet been reported.

An example of a calibration method that can be employed for the biochip temperature sensor involves placing the biochip in a constant-temperature water bath and measuring the resistance and voltage of the temperature sensor against fundamental temperatures in each application. However, in the case involving the constant-temperature water bath,

<sup>\*</sup> Corresponding Author

there is a problem with the long calibration time required by the biochip temperature sensor as the temperature of the constant-temperature water bath changes slowly. Therefore, in this paper, we propose a calibration system and a method that calibrates the biochip temperature sensor by measuring the surface temperature of the biochip using a biochip temperature sensor and a precision temperature sensor.

# 2. Methods



Figure 1. Block Diagram of Biochip Calibration System and QR Code Recognition System

Figure. 1 is a block diagram of the biochip calibration system and the quick response (QR) code recognition system. The biochip driving mechanism has a microprocessor (local system processor), and it controls or monitors each component of the biochip via a peripheral interface. For example, the temperature sensor is converted to a digital sensor using the analog-to-digital converter (ADC) of the microprocessor. The electric power for the heater is controlled through a pulse width modulation (PWM) device. The polymerase chain reaction (PCR) chip driving mechanism is connected to the PC using a USB interface. The PCR driving mechanism and the PCR chip interface are driven by distributing the USB power supply through power supply circuits within the driving mechanism.

The speed of the biochip used in this paper is measured as  $10^{\circ}$ C per second as it reaches 95 °C from the room temperature. The temperature is measured by heating the biochip and shifting to a temperature range that is important for each application. Then,

the ADC value is sent to the PC from the thermistor that is attached to the biochip, and in the process passing the biochip driving mechanism. The precision measurement device measures the biochip temperature at an important temperature for each application as it is attached to the surface of the biochip. The measured value is sent to the PC through the precision sensor driving mechanism. The PC compares the biochip temperature sensor value and the temperature value that has been sent from the precision measuring sensor, and it records the value of the calibration factor. The value of the calibration factor can be stored or recorded using various methods, which include saving it as a file or saving it as a OR code. This paper uses the OR code because embedding the calibration value in each biochip significantly increases user convenience. This is because a different calibration value should be applied for each biochip, which contains the corrected information, when a user uses the biochip. To store the calibration value on the biochip, both ROM and flash memory can be used, but they are much more expensive than using the OR code printed on paper. The PC prints out the calibration value on the paper using the QR code, and the printed OR code is attached to the biochip. When the user uses this biochip, a biochip controller uses a webcam (SDC-100) to capture the attached QR code, and the captured images are sent to the PC. Then, the image delivered to the PC is converted to a calibration parameter by the QR code decoder.



Figure 2. Precision Sensor Attached to the Biochip Surface

Figure. 2 shows the precision sensor attached to the surface of the biochip using an aluminum tape to calibrate the biochip. Here, the aluminum tape is used because it is easy to transfer the temperature. When the ADC value is sent to the PC from the thermistor attached to the biochip, using the biochip driving mechanism, Eq. (1) is used to calculate the temperature value.

$$T^{-1} = A + B \cdot \ln R + C \cdot (\ln R)^3$$
(1)

In Eq. (1), A, B, and C are Steinhart–Hart coefficients, and R represents the measured resistance value of the thermistor. To solve for the Steinhart–Hart coefficient that converts to the temperature, we require the temperatures of the three sections. To set the temperatures for the three sections, we apply the temperature used in the process that amplifies the DNA. A 3-step process and a 2-step process are usually used to amplify the DNA. In the case of the 3-step process, sections of 92–96°C (denaturing),  $50-65^{\circ}$ C (primer annealing), and  $-72^{\circ}$ C (primer extension) are used, and in the case of the 2-step process, sections of  $92-96^{\circ}$ C (primer annealing) are used.

Here, the Steinhart–Hart coefficient is solved using the temperatures of the three sections (60, 72, 95) to calibrate the temperature sensor. The Steinhart–Hart coefficient can be solved using a cubic equation after substituting the standard temperatures provided in the data sheet (T1, T2, T3) and resistances corresponding to the standard temperatures (R1, R2, R3) in Eq. (1).



Figure 3. GUI of Biochip Calibration System

Figure. 3 illustrates the GUI of the biochip calibration system. The value of the biochip temperature shown in C1 is the temperature value calculated by applying the resistance value delivered to the USB and the Steinhart–Hart coefficient value in Eq. (1). C2 indicates the progress of the chip calibration. C3 automatically records the difference between the biochip temperature value and the value of the precision temperature sensor as the processes 1 to 3 in C2 are performed. C4 represents the changed Steinhart–Hart coefficient value. The value of C4 solves for (A', B', C') using Eq. (1) with resistance values found by Eq. (2), as follows.

$$R = \exp\left(\sqrt[3]{\beta - \alpha} - \sqrt[3]{\beta + \alpha}\right),$$
where  $\alpha = \frac{A - \frac{1}{T}}{2C}$  and  $\beta = \sqrt{\left(\frac{B}{3C}\right)^3 + \alpha^2}$ . (2)

Upon completion of the calibration, it is finally saved as the Steinhart–Hart coefficient, and it is printed out as the QR code. The QR code output is attached to the biochip surface.



Figure 4. Flow Chart for Biochip QR Code Recognition

Figure. 4 is a flow chart that recognizes the QR code attached to the biochip through a webcam. When the program starts, it waits until the biochip is recognized. If the biochip is recognized in the device, the video camera preview is operated. The preview recognizes the QR code. Then, if the number of QR code recognition attempts is exceeded, a warning window is outputted. The recognized QR code is re-calculated using the Steinhart–Hart coefficient value that was analyzed by the decoder, and is expressed on the program.



Figure 5. GUI of Biochip Calibration Check Program

Figure. 5 is the GUI for the calibration program that recognizes the QR code attached to the biochip using the webcam. The image recognized by the webcam outputs the Steinhart–Hart coefficient using the QR code decoder. After substituting into Eq. (1) using the output value, the temperature value is recalculated and expressed on the screen.

## **3.** Calibration Accuracy Evaluation

To evaluate the precision of the calibration system before the calibration, the same basic Steinhart–Hart coefficient was used. When the basic Steinhart–Hart coefficient is used, there was an error of  $\pm 0.4$  °C. Table 1 shows the data that were measured using the basic Steinhart–Hart coefficient, while Table 2 shows the data that were measured using the Steinhart–Hart coefficient calibrated for each chip. The maximum error for the temperature measurement in Table 1 is 0.4 °C, and the average error for the temperature measurement is about 0.161 °C. On the other hand, the maximum error for the temperature measurement in Table 2 is 0.1 °C, and the average error for the temperature measurement is 0.037 °C. Therefore, the proposed calibration system improves the accuracy and reduces the calibration time.

| Reference Temperature | 60.0°C | 72.0°C | 95.0°C |
|-----------------------|--------|--------|--------|
| Thermistor no. 1      | 60.2°C | 72.0°C | 95.1°C |
| Thermistor no. 2      | 60.2°C | 72.0°C | 95.2°C |
| Thermistor no. 3      | 60.2°C | 71.9°C | 95.2°C |
| Thermistor no. 4      | 60.2°C | 71.7°C | 95.2°C |
| Thermistor no. 5      | 60.2°C | 72.0°C | 95.1°C |
| Thermistor no. 6      | 60.2°C | 72.0°C | 95.1°C |
| Thermistor no. 7      | 60.2°C | 72.0°C | 95.1°C |
| Thermistor no. 8      | 60.2°C | 72.0°C | 95.1°C |
| Thermistor no. 9      | 60.0°C | 72.3°C | 94.9°C |
| Thermistor no. 10     | 60.1°C | 72.4°C | 95.0°C |
| Thermistor no. 11     | 60.0°C | 72.3°C | 94.8°C |
| Thermistor no. 12     | 60.1°C | 72.4°C | 95.0°C |
| Thermistor no. 13     | 60.2°C | 72.1°C | 94.9°C |
| Thermistor no. 14     | 60.2°C | 71.9°C | 95.0°C |
| Thermistor no. 15     | 60.0°C | 72.2°C | 94.9°C |
| Thermistor no. 16     | 60.0°C | 72.2°C | 94.9°C |

Table 1. Measured Temperature before the Calibration (°C)

### 4. Conclusion

In this paper, we proposed a system that calibrates the temperature sensor of the biochip by directly heating the biochip and using the difference between the measured values of the temperature sensor attached to the biochip and the precision temperature sensor on the biochip surface. In addition, directly heating the biochip was observed to shorten the time taken to reach the target temperature, compared to the chip calibration by placing it in the constant-temperature water bath. The shortened calibration time of the biochip enables us to realize reduced production cost when the chips are manufactured on a large scale.

| Reference Temperature | <b>60.0</b> ℃  | <b>72.0℃</b> | <b>95.0</b> ℃ |
|-----------------------|----------------|--------------|---------------|
| Thermistor no. 1      | <b>60.0</b> ℃  | <b>72.0℃</b> | <b>95.1</b> ℃ |
| Thermistor no. 2      | 60.0°C         | <b>72.0℃</b> | <b>95.0</b> ℃ |
| Thermistor no. 3      | 60.0°C         | 71.9°C       | <b>95.0</b> ℃ |
| Thermistor no. 4      | <b>60.0</b> °С | 71.9°C       | <b>95.0</b> ℃ |
| Thermistor no. 5      | <b>60.0</b> °С | <b>72.0℃</b> | <b>95.1</b> ℃ |

Table 2. Measured Temperature after the Calibration (°C)

International Journal of Bio-Science and Bio-Technology Vol.7, No.2 (2015)

| Thermistor no. 6  | <b>60.0</b> ℃  | <b>72.0</b> ℃ | <b>95.1</b> ℃ |
|-------------------|----------------|---------------|---------------|
| Thermistor no. 7  | <b>60.0</b> °С | <b>72.0℃</b>  | <b>95.1</b> ℃ |
| Thermistor no. 8  | <b>60.0</b> °С | <b>72.0℃</b>  | <b>95.1</b> ℃ |
| Thermistor no. 9  | 60.1 °C        | 72.1 °C       | 95.0℃         |
| Thermistor no. 10 | <b>60.0</b> °С | 72.1 °C       | 95.0℃         |
| Thermistor no. 11 | 60.1 °C        | <b>72.0℃</b>  | 95.0℃         |
| Thermistor no. 12 | <b>60.0</b> °С | 72.1 °C       | 95.0℃         |
| Thermistor no. 13 | <b>60.0</b> °С | 72.1 °C       | <b>94.9℃</b>  |
| Thermistor no. 14 | <b>60.0</b> °С | 71.1 °C       | 95.0℃         |
| Thermistor no. 15 | <b>60.0</b> °С | <b>72.1</b> ℃ | <b>94.9℃</b>  |
| Thermistor no. 16 | <b>60.0</b> ℃  | <b>72.1</b> ℃ | 94.9℃         |

### Acknowledgments

The research was supported by Basic Science Research Program through the National Research Foundation of Korea (NRF) funded by the Ministry of Education, Science and Technology (2012R1A1A2040381) and by the "Research and Business Development, 2013" project of Ministry of Trade, Industry and Energy (N0000907).

### References

- J. M. Bartlett and D. Stirling, "A short history of the polymerase chain reaction", PCR protocols, (2003), pp. 3-6.
   K. B. Mullis, H. A. Erlich, N. Arnheim, G. T. Horn, R. K. Saiki and S. J. Scharf, "One of the first Polymerase Chain Reaction (PCR) patents", Google Patents, (1987).
   R. K. Saiki, S. Scharf, F. Faloona, K. B. Mullis, G. T. Horn, H. A. Erlich and N. Arnheim, "Enzymatic amplification of beta-globin genomic sequences and restriction site analysis for diagnosis of sickle cell anemia", Science, vol. 230, (1985), pp.1350-1354.
   R. K. Saiki, D. H. Gelfand, S. Stoffel, S. J. Scharf, R. Higuchi, G. T. Horn, K. B. Mullis and H. A. Erlich, "Primer-directed enzymatic amplification of DNA with a thermostable DNA polymerase", Science, vol. 239, (1988), pp. 487-491.
   D. Consulting, "BioChips & BioMEMS—Global Markets", Applications & Competitors: Analysis & Forecasts, (2005).
- [6] J. G. Lee, K. H. Cheong, N. Huh, S. Kim, J. W. Choi and C. Ko, "Microchip-based one step DNA extraction and real-time PCR in one chamber for rapid pathogen identification", Lab on a Chip, vol. 6, (2006), pp. 886-895.
- [7] E. Salm, Y. S. Liu, D. Marchwiany, D. Morisette, Y. He, A. K. Bhunia and R. Bashir, "Electrical detection of dsDNA and polymerase chain reaction amplification", Biomedical microdevices, vol. 13, (2011), pp. 973-982.
- [8] J. Wu, R. Kodzius, K. Xiao, J. Qin and W. Wen, "Fast detection of genetic information by an optimized PCR in an interchangeable chip", Biomedical microdevices, vol. 14, (2012), pp. 179-186.
- [9] C. Koo, M. M. Wight, H. S. Kim, O. S. Cifci, V. L. V. Diaz, B. Ma, S. Kim, H. Abdel-Raziq, K. Ong and Y. K. Jo, "Development of a Real-Time Microchip PCR System for Portable Plant Disease Diagnosis", PloS one, vol. 8, (2013), pp. 82704.
- [10] C. Y. Park, J. D. Kim, J. H. Ku, Y. S. Kim, H. J. Song and J. Kim, "Printed Circuit Board-Based Polymerase Chain Reaction Chip, Sensor Letters", vol. 10, (2012), pp. 1197-1202.

### Authors



**Sang-Yoon Kim**, he received a master's degree in Computer Engineering from Hallym University. He is getting a PhD in Computer Engineering at Hallym University. His research interests are in Bio-IT convergence, embedded system



**Jong-Dae Kim**, he received the M.S. and the Ph.D. degrees in Electrical Engineering from Korea Advanced Institute of Science and Technology, Seoul, Korea, in 1984 and 1990, respectively. He worked for Samsung Electronics from 1988 to 2000 as an electrical engineer. He is a Professor in Department of Ubiquitous Computing, Hallym University. His recent interests focus on biomedical system and bioinformatics.



**Yu-Seop Kim,** he received the Ph.D. degree in Computer Engineering from Seoul National University. He is currently a Professor in the Department of Ubiquitous Computing at Hallym University, South Korea. His research interests are in the areas of bioinformatics, computational intelligence and natural language processing.



**Hye-Jeong Song**, she received the Ph.D. degree in Computer Engineering from Hallym University. She is a Professor in Department of Ubiquitous Computing, Hallym University. Her recent interests focuses on biomedical system and bioinformatics



**Chan-Young Park**, he received the B.S. from Seoul National University and the M.S. and the Ph.D. degree from Korea Advanced Institute of Science and Technology in 1989 and 1995, respectively. From 1991 to 1999, he worked at Samsung Electronics. He is currently a Professor in the Department of Ubiquitous Computing of Hallym University, Korea. His research interests are in Bio-IT convergence, Intelligent Transportation System and sensor networks