Extraction and Fractionation of Phospholipids from the Waste of Jambal Siam (*Pangasius Hypophtalmus*) Processing

Mirna Ilza^{*} and Dian Iriani

Faculty of Fisheries and Marine Science Universitas Riau, Indonesia mirna.ilza@yahoo.co.id, dhian.iriani@gmail.com

Abstract

This research aimed to extract and fractionate the waste of Jambal Siam (Pangasius hypophtalmus) fish processing with acetone and ethanol and also the characteristics of fatty acid of phospholipids fraction was yielded. The fractination process with acetone and ethanol estimated that the fractination of Phospholipids with different composition of fatty acid could yielded. The result of this research indicate that the highest saturated fatty acid contents on waste processing jambal siam was palmitate acid which estimated predominante in phosphatidylglycerol (PG), phosphatidic (PA), and kardiolipin (DPG) forming. The highest of unsaturated fatty acid contents was oleat acid which estimated predominate in phosphatidylinositol (PI), phosphatidyl ethanolamine (PE), and phosphatidylcholine (PC) forming.

Keywords: acetone, ethanol, Pangasius hypophtalmus, waste

1. Introduction

1.1. Background

Jambal siam (*Pangasius hypophtalmus*) fish or better known by the public as catfish that live in pond and cage, is one of many farmed fish are cultivated by the people in Riau province, especially in Kampar regency. According to [1] the production of farming jambal siam fish in 2008 reached 13.206 tons, this production increased from 2007 with total production of 6.391 tons, and in 2006 the total production of 3.394 tons. In 2011 Kampar regency can increase the production of jambal siam fish about 50 tons per day, in 2015 the production has reached 100 tons per day. This jambal siam fish is a fish that many consumers preferred in fresh condition and has a high fat content relatively.

Based on these data, it can be seen that the farming of jambal siam fish in Riau province continues to increase every year so that it can lead to an abundance production of jambal siam fish. One effort to overcome the abundance of production and saturation of the consumer to fresh jambal siam fish is by doing business of fishery products processing. Jambal siam fish processing conventionally that many people do is the smoked fish and salted fish, while non conventional has not been done. Processing will produce waste in the form of fish entrails are intended to be discarded and not used. In addition to waste from processing, waste fish entrails jambal siam also came from the rest of the cuts of fish for household consumption and industrial restaurants in Pekanbaru and surrounding areas. The relatively large amount of waste, especially when coupled with the waste of other fish species that are also widely cultivated in Kampar district particularly and Riau Province generally.

Utilization of fish jambal conjoined so far is limited only as food. This gives the consequences of the economic value of fish jambal siam relatively low. To increase the economic value of fish jambal siam necessary research leading to production of non-food products of superior, one of which is the raw material drug industry. These opportunities

can be opened ranging from belly fat fish waste jambal siam, suspected to contain polyunsaturated fatty acids are relatively high. Several previous studies have shown that some parts, the abdominal fat and the liver often discarded for some reason when the fish are prepared for consumption, in terms of nutritional value and health effects are good especially because the content of EPA and DHA. Valuable fish oil can be obtained from fish waste. Fish waste is usually composed of a head, bones and tissue engaging member that is rich in fat and protein. Since so many health benefits of fish oil for the potential jambal siam fish waste needs to be improved for the source of local fish oils have the opportunity to be used as raw material for the pharmaceutical industry.

Waste entrails jambal siam fish consists of the digestive tract, fat, liver, and other organs. In the belly of the jambal siam fish there are about 1-2% fat pale yellow colored and solid form. These fats are classified as lipids which is located near the digestive tract jambal siam fish. Fats from jambal siam fish will be more in line with the expanding body size of fish. In addition, the fat content jambal siam fish will also increase the fish mature gonad.

Phospholipid (glycerophospholipid) a class of lipid compounds and is part of the membrane of living cells. Part of fat that is important enough to be in the cell is a phospholipid that is fat-containing phosphor. Lecithin is an especially important phospholipid found in cell membranes. Phospholipid consist of four components: fatty acids, a phosphate group, an alcohol containing nitrogen, and a framework. Phospholipid have a framework of glycerol and 2 acyl groups. In the third position of the glycerol frame is occupied by a phosphate group to the amino alcohol.

Food materials like fish oil and egg yolk included several classes of phospholipids: phosphatidylcholine (PC), phosphatidylethanolamine (PE), sphingomyelin (SM), lysophosphatidylcholine (LPC), lysophosphatidylcholine (LPE), and phosphatidylglycerol (PG). The most important phospholipid fractions of fish oil are PC and PE. Phospholipids are analyzed by chromatographic techniques, such as the following: thin layer chromatography (TLC), gas chromatography (GC).

The demand for various types of phospholipid is current constantly increasing because of its use in the industry continues to increase. Phospholipid are used for food product, medicinal formula, stabilizer, lubricant, cosmetic, pharmaceutical ingredient, and as an emulsifier. Synthetic phospholipid can be used for food and medicine, but now the interest of consumers switching to phospholipid from natural material.

Phospholipids have functional properties to health, market demand often require phospholipids fatty acids such as omega 3 changes in fatty acid composition can be made through the hydrolysis reaction [2].

Omega-3 fatty acids in phospholipid structure is more stable to oxidation than in the form of free fatty acids or triglycerides [3]. Phospholipids containing omega-3 fatty acids can be obtained from fish. In jambal Siamese fish belly fat containing phospholipids are the rest of the processing is not used. Therefore it should be used to meet consumer needs for phospholipids containing omega 3. Gladkowski [4] revealed that changes the molecular structure of phospholipids is intended to obtain properties suitable phospholipids different functional properties of phospholipids origin. The new Fosfolipid with the resulting chemical properties can be obtained by changing the type of fatty acids in the phospholipids with an organic solvent.

Riau province has potential phospholipid sources at its optimum, ie phospholipid from fish processing waste jambal siam. Most phospholipids are still in the fish processing waste jambal siam. Assessment phospholipid from fish processing waste jambal siam is important.

The fatty fish consists of various types of triglyceride is a molecule composed of glycerol and fatty acid [5], and triglyceride soluble in acetone, but insoluble lecithin polar compound [6, 7]. For ease of handling highly viscous crude lecithin and to improve the

dispersibility in water, the industry usually use acetone. Acetone extraction causes the phospholipid become concentrated thus increasing the level.

The egg yolk phospholipid extracted by using ethanol, followed by the extraction of acetone to remove fat from ethanol fraction [8, 3]. In this study examines methods of extraction and fractionation of phospholipid from fish processing waste jambal siam with ethanol and acetone and characteristics phospholipid fraction produced. The process of fractionation with ethanol and acetone could be expected to produce a phospholipid fraction with different fatty acid composition.

1.2. Formulation of the Problem

Jambal siam fish processing into smoked fish and salted fish in Kampar regency produces waste in the form fish entrails are intended to be discarded and not used. The relatively large amount of waste, especially when coupled with the waste of other fish species that are also widely cultivated in Kampar regency especially and Riau Province generally.

Jambal siam fish processing waste consisting of the gastrointestinal tract, fat, liver, and other organs. In the belly of the jambal siam fish there are about 1-2% fat pale yellow colored and solid form. These fat are classified as lipid which is located near the fish digestive tract jambal conjoined. In the lipid contains phospholipid required by the food industry, medicine, pharmaceutical, and cosmetic.

Given the lack of research on the extraction and fractionation of fish processing waste jambal siam, therefore it is important to do research so that fish waste has added value. In addition, this study also important for those who need information about the extraction and fractionation of phospholipid jambal siam fish waste as well as meet the need of industrial to natural phospholipid.

1.3. Research Purposes

The purpose of this research was to extract and fractionate phospholipid from the waste jambal siam (*Pangasius hypophtalmus*) fish processing.

2. Research Methods

2.1. Location

This research was conducted at the Laboratory of Food Chemistry Department of Fishery Product Technology Faculty of Fisheries and Marine Sciences Universitas Riau, and the Integrated Laboratory of Bogor Agricultural Institute.

2.2. Materials and Tools

Raw materials used in this study was jambal siam fish waste (Figure 1). The fish waste derived from Kampar regency. The chemicals used were the standard mixture of fatty acid, methylenechloride, NaOH, methanol, chloroform, ethanol, acetone, distilled water, and nitrogen gas.



Figure 1. Jambal Siam Fish Waste

The equipment used was water bath shaker, gas chromatography, oven, UV lamp, glass tools, and a rotary evaporator. The study is primarily based on the extraction of total lipid, phospholipid separation of total lipid, phospholipid fractionation, and identify the constituent fatty acid.

2.3. Research Procedure

2.3.1. Extraction of Fat Fish

To determine the fat content of the fish was using Soxhlet method [9], with the following procedures:

- Samples were crushed (preferably dry and passing 40 mesh) and weighed as much as 50 g and put into a Soxhlet extraction tube in a paper lead.
- Cooling water flowed through the condenser.
- Test tube mounted on soxhlet distillation with diethyl ether solvent of 100 ml for 5 hours. Then repeated until the unused sample weight of 400 g (8 repetitions).
- The second and subsequent 50 ml diethyl ether was added to each sample repetitions.
- Diethyl ether containing extract fat that has been removed from the soxhlet and dried in an oven for 2 hours at a temperature of 60°C.
- Then cooled in a desiccator for 30 minutes and weighed. Drying in the oven forwarded to constant weight. Heavy residue in the weighing bottle was expressed as the weight of fatty fish (Figure 2).

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Figure 2. Fatty Fish

2.3.2. Separation of Phospolipids Crude Extract of Total Lipid by Dissolution

Total lipid obtained from the extraction process was extracted using chloroform [8]. A total of 10 g of total lipid extracted with 40 ml of chloroform. A chloroform-soluble fraction and a non-polar lipid were separated by centrifugation at a speed of 5000 rpm for 10 minutes. Insoluble fraction of a polar lipid and separated. Soluble fraction was dried using nitrogen/aeration and re-extracted using 30 ml of chloroform, and then centrifuged to separate soluble and insoluble fraction.

Insoluble fraction of a polar lipid and mixed with the insoluble fraction from the first extraction. Furthermore chloroform insoluble fraction extracted by using 20 ml of methanol to dissolve the polar phospholipid. Insoluble fraction was separated by centrifugation at 5000 rpm for 10 minutes. Soluble fraction was dried to take solid which were phospholipid (Figure 3 a, b)

2.3.3. Phospholipid Fractionation

Extracted phospolipid was dissolved in ethanol and acetone. The first fractionation was done by using ethanol. Ethanol soluble fraction further fractionated using acetone.

Fractionation process was performed as follows: 5 g phospholipid crude dissolved in 20 ml of ethanol, agitated for 60 minutes. Supernatant and the precipitate was separated by centrifugation at 5000 rpm for 10 minutes. The residue was taken and an insoluble fraction of ethanol. Ethanol soluble fraction was taken by evaporating ethanol by spraying nitrogen gas/air.

Ethanol soluble fraction was taken partly for and partly characterized further fractionated by dissolving acetone (1: 4 w/v) and agitated for 60 minutes. Acetone insoluble fraction was separated from the solution by centrifugation at 5000 rpm for 10 minutes (Figure 4 a, b).

Acetone soluble fraction was taken by evaporating acetone with nitrogen gas/air. The results of the four fractions obtained by fractionation of phospholipid that fraction insoluble ethanol, insoluble fraction of ethanol, ethanol and acetone soluble fraction, the

fraction insoluble ethanol and acetone, and phospholipid without fractionation or crude phospholipid. Fractions are then identified the types of constituent fatty acid [8].

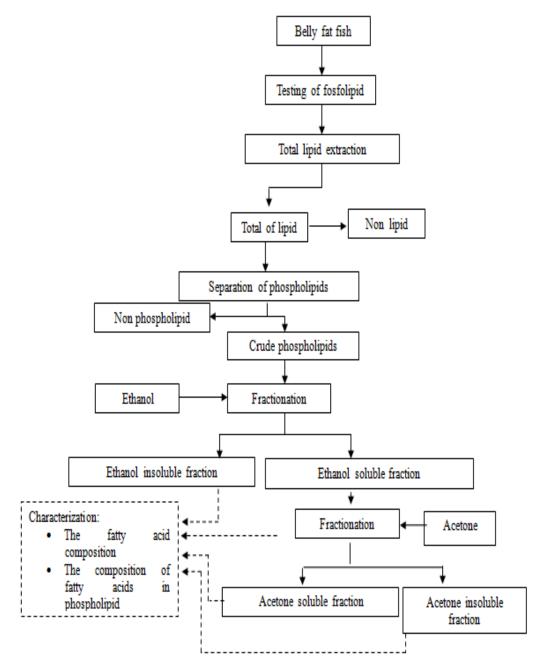


Figure 3 a. Total Lipid Extraction, Separation, and Fractionation of Phospholipids

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Figure 3 b. Phosfolipid

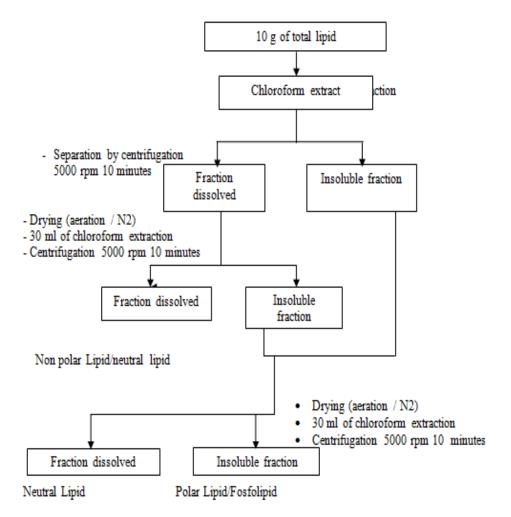


Figure 4 a. Phospholipids Separation of Total Lipids from the Solvent



Figure 4 b. Ethanol Soluble Fraction and Insoluble Fraction Aceton Soluble Fraction and Insoluble Fraction

2.3.4. Identify the Type of Fatty Acid in Crude Phospholipid and Phospholipid Fraction

Identify the type of fatty acid of any phospholipid fraction carried by gas chromatography [10]. The identification was done by comparing the retention time of the standard mixture of fatty acid that was injected separately. Quantification was done based on the relative percentage.

2.3.5. Data Analysis

The data obtained were processed descriptively and presented in tabular form. Further data on fractionation and fatty acids contained in each fraction discussed using crude phospholipid fraction as a comparison.

3. Results and Discussions

3.1. Saturated Fatty Acid (SAFA)

The results of the analysis of saturated fatty acids (Saturated Fatty Acid /SAFA) using gas chromatography showed that saturated fatty acids were detected in each of the different phospholipid fraction amount. In each of the fractions can be detected as many as 11 kinds of saturated fatty acids to total saturated fatty acids 35.94% - 44.27%.

	Amount (%)					
Fatty acid type	Crude fosfolipid	Soluble in ethanol	Insoluble in ethanol	Soluble In aceton	Insoluble in acetone	
Lauric acid, C12:0	1.94	1.26	1.50	118	1.30	
Myristic acid, C14:0	4.51	3.83	4.07	3.75	3.87	
Pentadekanoat acid, C15:0	1.99	1.31	1.55	1.23	1.35	
Acid Palmitate, C16:0	18.83	18.15	18.37	1.07	18.19	
Heptadekanoat acid, C17:0	2.02	1.34	1.58	1.26	1.38	
Stearic acid, C18:0	5.26	4.53	4.82	4.50	4.62	
Arakhidat acid, C20:0	2.08	1.40	1.64	1.32	1.44	
Heneikosanoat acid, C21:0	1.88	1.20	1.44	1.12	1.24	
Behenic acid, C22:0	1.93	1.27	1.51	1.19	1.31	
Trikosanoat acid, C23:0	1.89	1.21	1.45	1.13	1.25	
Lignoserat acid, C24:0	1.94	1.26	1.50	1.19	1.34	
Saturated Fatty Acid Total	44.27	36.76	39.43	35.94	37.29	

Table 1. Saturated Fatty Acid Composition of Phospholipid Fraction

Highest amount of fatty acid was palmitic acid, *i.e.*, on crude phospholipid 18.83%, ethanol soluble phospholipid 18.15%, phospholipid insoluble ethanol 18.37%, acetone insoluble phospholipid 18.07%, and phospholipid insoluble acetone 18.19%. For more details, the results of the analysis can be seen in Table 1.

The palmitic acid dominates the formation phospatidilgliserol (PG), phosphatidic acid (PA), and cardiolipin (DPG). Differences in the fatty acid composition of each fraction of phospholipids was expected to affect the functional properties of phospholipids such as emulsification ability [8].

3.2. Monounsaturated Fatty Acids (MUFA)

Monounsaturated fatty acids/ MUFA) fractionation of phospholipids were analyzed using gas chromatography was different amount. The type and amount of fatty acids contained in each fraction are shown in Table 2.

	Amount (%)					
Fatty acid type	Crude Fosfolipid	Soluble in Ethanol	Insoluble in ethanol	Soluble in acetone	Insoluble in acetone	
Myristoleic acid, C14:1	0.15	0.99	0.02	1.32	1.59	
Palmitoleic acid, C16:1	0.80	1.64	0.53	1.97	2.24	
Cis-10-Heptadekanoat	1.00	1.04	0.07	1.37	1.64	
Elaidic acid, C18:1n9t	0.23	1.07	0.10	1.40	1.67	
Oleic acid, C18:1n9c	19.64	21.31	16.34	2.,63	21.91	
Cis-11-Eikosenoat acid, C20:1	0.65	1.49	0.38	1.82	2.09	
Erucat acid, C22:1n9	0.16	1.02	0.03	1.33	1.61	
Mono Unsaturated Fatty Acid Total	22.63	28.56	17.47	30.84	32.75	

 Table 2. Mono Unsaturated Fatty Acids Composition of Phospholipid

 Fraction

Table 2 shows that in each of the fractions can be detected as many as seven types of monounsaturated fatty acids to total monounsaturated fatty acids 17.47% - 32.75%. The highest fatty acid was oleic acid, which was the crude phospholipid 19.64%, ethanol soluble phospholipids 21.31%, phospholipids insoluble ethanol 16.34%, acetone insoluble phospholipids 21.63%, and phospholipids insoluble acetone 21.91 %.

Ethanol soluble fraction had levels of monounsaturated fatty acids (oleic and palmitoleic) higher than the ethanol insoluble fraction. Acetone is a solvent semipolar nature so that dilution with acetone produce levels of unsaturated fatty acids were almost equally between fractions soluble and insoluble acetone. Differences in the fatty acid composition is affected by the solvent. The fatty acid have a single double bond tend to be polar so that more soluble in ethanol.

The fatty acids dominate for phosphatidylinositol (PI), phosphatidyl ethanolamine (PE), and phosphatidylcholine (PC) was oleic acid [8, 11-12]. Fractionation was intended to get phospholipid fractions desired. The soluble fraction of ethanol was expected to produce fractions with the ratio of PC/PE was the proper height for oil in water emulsion, while the insoluble fraction of ethanol yield ratio of PC/PE low suitable for water-in-oil emulsion. Rao [3] reported the lipid classes and fatty acid composition of eggs from the Atlantic halibut (*Hippoglossus hippoglossus*) were studied. The phospholipid fraction contained 62% PC and 7 % PE with high concentration of n-3 PUFA such as EPA and DHA.

3.3. Polyunsaturated Fatty Acid (PUFA)

Type of polyunsaturated fatty acid (Poly Unsaturated Fatty Acid / PUFA) contained in each fraction there are 11 types. For more details, type and amount of polyunsaturated fatty acid in fatty fish each can be seen in Table 3.

In Table 3 shown that the total of polyunsaturated fatty acid was 13.24% - 22.43%. The highest fatty acid was linoleic acid, which was the crude phospholipid 10.14%, ethanol soluble phospholipid 10.20%, ethanol insoluble phospholipid 9.41%, acetone insoluble phospholipid 9.43%, and phospholipid insoluble acetone 10.99%.

Ethanol soluble fraction had level of polyunsaturated fatty acid was higher than ethanol insoluble fraction. Ethanol was a polar solvent that was able to extract the fatty acids which its polarity level same with solvent. While acetone was a semipolar solvent that produce level of unsaturated fatty acid were almost equally between acetone soluble and

acetone insoluble fractions. In essence of a material to be easily dissolved in solvent with the same of polarity, because of fat polarities was different so there was no general solvent for all types of fat [13]. Therefore necessary fish oil extraction process in accordance with needs. These results indicate that the soluble fraction of ethanol can increase the concentration of omega-3.

Rao *et al.*, [14] reported EPA and DHA in the phospholipid fractions were 2.0, 7.9% in catla and 0.3 and 6.3% in mrigal respectively. Polyunsaturated fatty acids (EPA and DHA) were found to be more concentrated in the phospholipid fractions. EPA was found in smaller amounts while DHA was observed in significant amounts. The concentration of EPA and DHA in significant amounts in the phospholipid fraction was also observed in other roe lipids studied earlier. The fish roes of rohu and murrel showed the presence of eicosapentaenoic acid (20:5, EPA) to an extent of 1.1 and 1.0% and docosahexaenoic acid (22:6, DHA) to an extent of 14.1 and 6.8% respectively in the phospholipid fraction.

	Amount (%)						
Fatty acid type	Crude Fosfolipid	Soluble in ethanol	Insoluble in ethanol	Soluble in aceton	Insoluble in Aceton		
Linolelaidat asid, C18:2n9t	0.70	0.86	0.02	0.73	0.80		
Linoleic, C18:2n6c	10.14	10.20	9.41	9.43	10.99		
G-Linolenic acid, C18:3n6	1.12	1.23	0.39	1.10	1.17		
Linolenic acid, C18:3n3	1.18	1.29	0.45	1.16	1.23		
Cis-11,14- Eikosedienoat acid, C20:2	1.16	1.27	0.43	1.14	1.21		
Cis-8,11,14- Eikosetrienoat acid, C20:3n6	1.44	1.55	0.71	1.42	1.49		
Cis-11,14,17- Eikosetrienoat acid, C20:3n3	0.77	0.88	0.04	0.75	0.82		
Arachidonic acid,C20:4n6	1.35	1.46	0.62	1.33	1.43		
Cis-13,16- Dokosadienoat acid, C22:2	0.75	0.86	0.02	0.74	0.80		
Cis-5,8,11,14,17- Eikosapentaenoat acid, C20:5n3	1.01	1.12	0.28	1.72	1.26		
Cis- 4,7,10,13,16,19- Dokosaheksaenoat acid, C22:6n3	1.60	1.71	0.87	158	1.65		
Poly Unsaturated acid total	21.22	22.43	13.24	21.10	21.85		

Table 3. Poly Unsaturated Fatty Acids Composition Phospholipid Fraction

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The fatty acid profile of total lipids showed that the saturated fatty acids were found to an extent of 52.0 and 52.8% with major fatty acid being hexadecanoic acid to an extent of 34.7 and 37.5% in catla and mrigal eggs respectively. However, the saturated octadecanoic (13.8, 12.3%) and monounsaturated octadecenoic acid (22.9, 17.9%) were also present in considerable quantities. The octadecadienoic acid and octadecatrienoic acids were 3.3, 2.0% and 2.9, 6.0% in catla and mrigal lipids respectively. EPA (20:5) and DHA (22:6) were found to be 2.0, 6.9% and 0.5, 8.9% in catla and mrigal roe lipids respectively [3].

4. Conclusion

The research concluded that the highest of saturated fatty acid level was found in waste of jambal siam fish processing was palmitic acid which was suspected dominates the formation phospatidylgliserol (PG), phosphatidic acid (PA), and cardiolipin (DPG). The highest of unsaturated fatty acid level was oleic acid which allegedly dominate the formation of phosphatidylinositol (PI), phosphatidyl ethanolamine (PE), and phosphatidylcholine (PC).

Ethanol was a polar solvent that produced the fraction of unsaturated fatty acid of soluble ethanol higher than insoluble ethanol. While acetone was a solvent that was semipolar so that dilution with acetone produced level of unsaturated fatty acid were almost equally between aceton soluble fraction and acetone insoluble fraction.

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Authors



Mirna IIza obtained her Bachelor Degree (Ir) from Andalas University, Master Degree (M.S) degree from Padjadjaran University, and Ph.D degree from Andalas University, Indonesia. She is a professor at Department of Fishery Product Processing Faculty of Fisheries and Marine Universitas Riau Indonesia.



Dian Iriani obtained her Master degree (MP) from Brawijaya University in Malang East Java Indonesia and (M.Sc) degree from Burapha University Thailand. She is a lecturer in Department of Fishery Product Processing Faculty of Fisheries and Marine Universitas Riau Indonesia. International Journal of Bio-Science and Bio-Technology Vol.9, No.2 (2017)